# DATA NOTE

# Genome sequence of two novel virulent clinical strains of *Burkholderia pseudomallei* isolated from acute melioidosis cases imported to Israel from India and Thailand

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# Abstract

**Objective** *Burkholderia pseudomallei*, the etiological cause of melioidosis, is a soil saprophyte endemic in South-East Asia, where it constitutes a public health concern of high-priority. Melioidosis cases are sporadically identified in nonendemic areas, usually associated with travelers or import of goods from endemic regions. Due to extensive intercontinental traveling and the anticipated climate change-associated alterations of the soil bacterial flora, there is an increasing concern for inadvertent establishment of novel endemic areas, which may expand the global burden of melioidosis. Rapid diagnosis, isolation and characterization of *B. pseudomallei* isolates is therefore of utmost importance particularly in non-endemic locations.

**Data description** We report the genome sequences of two novel clinical isolates (MWH2021 and MST2022) of *B. pseudomallei* identified in distinct acute cases of melioidosis diagnosed in two individuals arriving to Israel from India and Thailand, respectively. The data includes preliminary genetic analysis of the genomes determining their phylogenetic classification in rapport to the genomes of 131 *B. pseudomallei* strains documented in the NCBI database. Inspection of the genomic data revealed the presence or absence of loci encoding for several documented virulence determinants involved in the molecular pathogenesis of melioidosis. Virulence analysis in murine models of acute or chronic melioidosis established that both strains belong to the highly virulent class of *B. pseudomallei*.

Keywords Burkholderia pseudomallei, Melioidosis, Genome sequence, Virulence genes, Phylogenetic tree

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### Objective

The Gram-negative bacterial pathogen Burkholderia pseudomallei is the etiological cause of melioidosis. As of 2018 there were 165,000 global estimated annual morbidities, including 89,000 deaths, mostly in Southeast Asia where the disease is endemic [1]. B. pseudomallei is considered a high priority biohazard due to its prevalence in soil and water, high virulence associated with inhalation exposure, low infective dose, high mortality rates, native resistance to a wide range of antibiotics, and paucity of an efficient licensed vaccine [2–4]. Melioidosis symptoms are nonspecific, thus hindering identification of the disease, which may inadvertently be diagnosed as tuberculosis or a common form of pneumonia [1, 5, 6]. The B. *pseudomallei* genome is highly plastic, resulting in significant sequence variability amongst strains [7]. Melioidosis cases are sporadically identified in nonendemic areas, usually associated with travelers or transport of goods from endemic regions. Due to extensive intercontinental traveling and the anticipated climate change-associated alterations of the soil bacterial flora, there is an increasing concern for inadvertent establishment of novel endemic areas which may expand the global burden of melioidosis. Rapid diagnosis, isolation and characterization of *B*. pseudomallei isolates is therefore of utmost importance, in particular in non-endemic locations.

Two Israelis returning from Southeast Asia, one from Thailand and the other from India, were hospitalized in different medical centers due to fulminant symptoms reminiscent of acute pneumonia, subsequently diagnosed as travel-related melioidosis. The objective of the study documented in this brief data note, was genetic identification of the etiologic pathogen isolated from these clinical cases, and analysis of the respective bacterial genomes for taxonomic typing and primary determination of their genetic characteristics.

#### **Data description**

We report the draft genome of two novel clinical *B. pseudomallei* strains denoted MWH2021 and MST2022 isolated from melioidosis cases diagnosed in Israel, associated with travelers from India and Thailand, respectively.

Previously, three additional clinical isolates of melioidosis were documented in Israel, from Thailand [8], Eritrea [9] and India [10, 11]. The strains described in the current report were acknowledged as *B. pseudomallei* by the Vitek KL2 bacterial identification system and MALDI-TOF (Bruker) mass spectrometric analysis and confirmed by PCR analysis [12]. Virulence analysis of the strains was performed in the melioidosis murine model [13, 14]. MWH2021 strain exhibited an intranasal (IN) Lethal Dose 50% (LD<sub>50</sub>; more than 30 days survival of mice inoculated with increasing doses, calculated by linear regression using the GraphPad Prism V5 software [10, 13, 15]) of 126 CFU and 950 CFU in the BALB/c and C57BL/6J strains of mice, respectively. LD<sub>50</sub> of strain MST2022 was 13 CFU and 83 CFU (BALB/c and C57BL/6J mice, respectively). Of note, Balb/c serves for modeling the acute form of melioidosis while C57BL/6J recapitulates the chronic form. The data indicate that both novel clinical isolates belong to the group of highly virulent strains [16].

Genome sequencing *of B. pseudomallei* MWH2021 and MST2022 isolates employed purified chromosomal DNA isolated from BHI-agar colonies, which served for generation of genomic libraries (Nextera XT kit, Illumina). Sequencing of both strains was performed on Miseq Instrument (Illumina), generating short-read sequences. Long-read sequencing using Oxford Nanopore Technologies, produced additional data sets which enabled improved assembly [13].

The genomic sequences of the two novel strains were deposited to the NCBI database [17-20]. The data deposited in the various public databases as well as supplementary material pertaining to this data note are detailed in Table 1. To determine the phylogenetic relation of the novel strains to other B. pseudomallei strains. A total of 131 complete genome sequences were downloaded from NCBI [21]. Core genome alignment and phylogeny of the strains relative to the reference genome strain Mahidol-1106a (accession number GCF\_000756125.1) were performed using the Parsnp software, v1.2 [22]. The resulting phylogenetic tree, depicted in the Supplementary Fig. 1 [23] was shaped with the iTOL: interactive tree of life platform [24]. This analysis established that the novel MWH2021 strain is adjacent in the phylogenetic clustering to strain MAA2018 [10, 11], in line with their common geographic origin (India). The two isolates differ by 8,879 and 7,408 SNPs on chromosome 1 and 2, respectively, clearly indicating that in spite of their phylogenetic vicinity they represent distinct strains. In general, as expected, the analysis established a strong correlation between genomic DNA sequence similarities of various strains and their respective geographical origin (see Supplementary Fig. 1).

The genomes of the novel strains, as well as those of the three additional strains BP1, BP2 and MAA2018 previously isolated and documented in Israel from cases of melioidosis [8–11, 13], were interrogated for the presence of 36 genes encoding for potential *B. pseudomallei* virulence factors [1, 3]. The results summarized in Supplementary Data 2 [14], show that all these genes are present in the genome of the novel strain MWH2021. In this regard, this strain does not differ from the previously documented strain of Indian origin, in line with their phylogenetic proximity. Three genes (*chbP*, *boaB* and *boaA*) were not identified in the genome of the novel

#### Table 1 Overview of data files/data sets

| Label                                   | Name of<br>data file/<br>data set   | File types<br>(file<br>extension)   | Data repository and<br>identifier (DOI or ac-<br>cession number) | Ref.<br>(Repos-<br>itory) |
|---|---|-------------------------------------|--|---------------------------|
| Data<br>set 1                           | <i>B. pseu-<br/>domallei</i><br>MWH2021<br>assembly                               | DNA<br>sequence<br>(.fasta)         | https://identifiers.<br>org/ncbi/insdc.<br>gca:GCA_030913145.1   | 17<br>(NCBI)              |
| Data<br>set 2                           | <i>B. pseu-<br/>domallei</i><br>MST2022<br>assembly                               | DNA<br>sequence<br>(.fasta)         | http://identi-<br>fiers.org/insdc.<br>gca:GCA_030144945.1        | 18<br>(NCBI)              |
| Data<br>set 3                           | B. pseu-<br>domallei<br>MWH2021<br>raw reads                                      | DNA<br>sequence<br>(.fastq)         | https://identifiers.<br>org/ncbi/insdc.<br>sra:SRP483724         | 19<br>(NCBI)              |
| Data<br>set 4                           | B. pseu-<br>domallei<br>MST2022<br>raw reads                                      | DNA<br>sequence<br>(.fastq)         | https://identifiers.<br>org/ncbi/insdc.<br>sra:SRP483736         | 20<br>(NCBI)              |
| Sup-<br>ple-<br>men-<br>tary<br>Fig. 1  | Phylo-<br>genetic<br>tree of <i>B.</i><br><i>pseudomal-</i><br><i>lei</i> strains | Microsoft<br>Power Point<br>(.pptx) | https://doi.<br>org/10.6084/<br>m9.figshare.24961800             | 23<br>(Fig-<br>share)     |
| Sup-<br>ple-<br>men-<br>tary<br>Data 1  | Supple-<br>mentary<br>Method<br>Data  | Microsoft-<br>Word<br>(.docx)       | https://doi.<br>org/10.6084/<br>m9.figshare.25204352             | 13<br>(Fig-<br>share)     |
| Sup-<br>ple-<br>men-<br>tary<br>Data 2  | LD <sub>50</sub><br>values and<br>virulence<br>determi-<br>nants                  | Microsoft-<br>Word<br>(.docx)       | https://doi.<br>org/10.6084/<br>m9.figshare.25204568             | 14<br>(Fig-<br>share)     |
| Sup-<br>ple-<br>men-<br>tary<br>Table 1 | NCBI <i>BP</i><br>complete<br>genomes<br>as of June<br>2023                       | Excel (.xls)                        | https://doi.<br>org/10.6084/<br>m9.figshare.25479535             | 28<br>(Fig-<br>share)     |

strain MST2022. While these genes encode for factors believed to be involved in the virulence of the bacteria [25–27], their absence did not correlate with decreased virulence of the MST2022 [14], in accordance with the notion that pathology of melioidosis involves the activity of numerous bacterial factors whose individual contribution to virulence is difficult to assess.

# Limitation

Bioinformatic DNA similarity analysis of the genomic sequences of the two novel clinical isolates, for determining their phylogenetic relation to other *B. pseudom-allei* strains [23, 28], was conducted using 131 complete *B. pseudomallei* genomes present in the NCBI databank (as of June 2023). It is conceivable that conducting the analysis by comparison to all available *B. pseudomallei* genomes (not only complete ones), may have provided a more comprehensive phylogenetic profile.

The bioinformatics screen of the sequences for potential virulence factors did not include inspection of the possible point mutations or indels (insertions/deletions) but only presence or absence of the respective orthologous genes, therefore the study cannot attest for their level of expression.

#### Abbreviations

| B. pseudomallei | Burkholderia pseudomallei                                |
|-----------------|--|
| MWH2021         | Melioidosis case from Wolfson Hospital, Holon, 2021      |
| MST2022         | Melioidosis case from Sheba Hospital, Tel-Hashomer, 2022 |

# **Supplementary Information**

The online version contains supplementary material available at https://doi. org/10.1186/s12863-024-01225-x.

| Supplementary Material 1 |  |
|--------------------------|--|
| Supplementary Material 2 |  |
| Supplementary Material 3 |  |
| Supplementary Material 4 |  |

#### Author contributions

TC, IC-G and OI wrote the manuscript; SA, IZ, OS and YM isolated the pathogens; OI, SL and AB-D performed the sequencing of the genomes; TC, MI, DG, MA, EB-H and UE performed the virulence experiments; IC-G, GB, GZ, AZ performed the bioinformatics analyses; OC, EM and TC supervised the project.

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#### Data availability

The data described in this Data note can be freely and openly accessed on NCBI under [GCF\_030913145.1; GCA\_030144945.1; PRJNA1008066; PRJNA962450]. Please see Table 1 and references [13, 14, 17-20, 23, 28] for details and links to the data.

## Declarations

**Ethics approval and consent to participate** Not applicable.

**Consent for publication** Not applicable.

# Competing interests

The authors declare no competing interest

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#### References

- Wiersinga WJ, Virk HS, Torres AG, Currie BJ, Peacock SJ, Dance DAB, Limmathurotsakul D, Melioidosis. Nat Rev Dis Primers. 2018;4:17107. https://doi. org/10.1038/nrdp.2017.107. PMID: 29388572; PMCID: PMC6456913.
- Limmathurotsakul D, Golding N, Dance DA, Messina JP, Pigott DM, Moyes CL, Rolim DB, Bertherat E, Day NP, Peacock SJ, Hay SI. Predicted global distribution of *Burkholderia pseudomallei* and burden of melioidosis. Nat Microbiol. 2016;1:15008. https://doi.org/10.1038/nmicrobiol.2015.8. PMID: 27571754.
- Meumann EM, Limmathurotsakul D, Dunachie SJ, Wiersinga WJ, Currie BJ. Burkholderia pseudomallei and melioidosis. Nat Rev Microbiol. 2023 Oct 4. https://doi.org/10.1038/s41579-023-00972-5. Epub ahead of print. PMID: 37794173.

- Loveleena, Chaudhry R, Dhawan B. Melioidosis; the remarkable imitator: recent perspectives. J Assoc Physicians India. 2004;52:417–20. PMID: 15656034.
- Le Tohic S, Montana M, Koch L, Curti C, Vanelle P. A review of melioidosis cases imported into Europe. Eur J Clin Microbiol Infect Dis. 2019;1–14. https:// doi.org/10.1007/s10096-019-03548-5.
- Holden MT, Titball RW, Peacock SJ, Cerdeño-Tárraga AM, Atkins T, Crossman LC, Pitt T, Churcher C, Mungall K, Bentley SD, Sebaihia M, Thomson NR, Bason N, Beacham IR, Brooks K, Brown KA, Brown NF, Challis GL, Cherevach I, Chillingworth T, Cronin A, Crossett B, Davis P, DeShazer D, Feltwell T, Fraser A, Hance Z, Hauser H, Holroyd S, Jagels K, Keith KE, Maddison M, Moule S, Price C, Quail MA, Rabbinowitsch E, Rutherford K, Sanders M, Simmonds M, Songsivilai S, Stevens K, Tumapa S, Vesaratchavest M, Whitehead S, Yeats C, Barrell BG, Oyston PC, Parkhill J. Genomic plasticity of the causative agent of melioidosis, *Burkholderia pseudomallei*. Proc Natl Acad Sci U S A. 2004;101(39):14240–5. https://doi.org/10.1073/pnas.0403302101. Epub 2004 Sep 17. PMID: 15377794; PMCID: PMC521101.
- Almog Y, Yagel Y, Geffen Y, Yagupsky P. A Burkholderia pseudomallei infection Imported from Eritrea to Israel. Am J Trop Med Hyg. 2016;95(5):997–8. https:// doi.org/10.4269/ajtmh.16-0481. Epub 2016 Aug 29. PMID: 27573625; PMCID: PMC5094250.
- Cahn A, Koslowsky B, Nir-Paz R, Temper V, Hiller N, Karlinsky A, Gur I, Hidalgo-Grass C, Heyman SN, Moses AE, Block C. Imported melioidosis, Israel, 2008. Emerg Infect Dis. 2009;15(11):1809–11. https://doi.org/10.3201/ eid1511.090038. PMID: 19891871; PMCID: PMC2857218.
- Brosh-Nissimov T, Grupel D, Abuhasira S, Leskes H, Israeli M, Lazar S, Elia U, Israeli O, Beth-Din A, Bar-Haim E, Cohen-Gihon I, Zvi A, Cohen O, Chitlaru T. Case Report: Imported Melioidosis from Goa, India to Israel, 2018. Am J Trop Med Hyg. 2019;101(3):580–584. https://doi.org/10.4269/ajtmh.19-0303. Erratum in: Am J Trop Med Hyg. 2019;101(5):1187. PMID: 31287043; PMCID: PMC6726963.
- Israeli O, Cohen-Gihon I, Brosh-Nissimov T, Zvi A, Beth-Din A, Shifman O, Israeli M, Elia U, Lazar S, Bar-Haim E, Cohen O, Chitlaru T. Draft genome sequence of a rare Israeli clinical isolate of Burkholderia pseudomallei. Microbiol Resour Announc. 2019;8(19):e00281–19. https://doi.org/10.1128/MRA.00281-19. Erratum in: Microbiol Resour Announc. 2019;8(40): PMID: 31072902; PMCID: PMC6509527.
- Price EP, Dale JL, Cook JM, Sarovich DS, Seymour ML, Ginther JL, Kaufman EL, Beckstrom-Sternberg SM, Mayo M, Kaestli M, Glass MB, Gee JE, Wuthiekanun V, Warner JM, Baker A, Foster JT, Tan P, Tuanyok A, Limmathurotsakul D, Peacock SJ, Currie BJ, Wagner DM, Keim P, Pearson T. Development and validation of *Burkholderia pseudomallei*-specific real-time PCR assays for clinical, environmental or forensic detection applications. PLoS ONE. 2012;7(5):e37723. https://doi.org/10.1371/journal.pone.0037723. Epub 2012 May 18. PMID: 22624061; PMCID: PMC3356290.
- Cohen-Gihon I, Chitlaru T. 2024. Figshare. https://doi.org/10.6084/ m9.figshare.25204352.

- 14. Cohen-Gihon I, Chitlaru T. 2024. Figshare. https://doi.org/10.6084/ m9.figshare.25204568.
- Chitlaru T, Israeli M, Bar-Haim E, Elia U, Rotem S, Ehrlich S, Cohen O, Shafferman A. Next-generation Bacillus anthracis live attenuated spore vaccine based on the htrA(-) (high temperature requirement A) Sterne Strain. Sci Rep. 2016;6:18908. https://doi.org/10.1038/srep18908. PMID: 26732659; PMCID: PMC4702213.
- 16. Warawa JM. Evaluation of surrogate animal models of melioidosis. Front Microbiol. 2010;1:141. https://doi.org/10.3389/fmicb.2010.00141. PMID: 21772830; PMCID: PMC3109346.
- Cohen-Gihon I, Chitlaru T. NCBI. 2023. https://identifiers.org/ncbi/insdc. gca:GCA\_030913145.1.
- Cohen-Gihon I, Chitlaru T. 2023. NCBI. http://identifiers.org/insdc. gca:GCA\_030144945.1.
- Cohen-Gihon I, Chitlaru T. 2023. NCBI. https://identifiers.org/ncbi/insdc. sra:SRP483724.
- Cohen-Gihon I, Chitlaru T. 2023. NCBI. https://identifiers.org/ncbi/insdc. sra:SRP483736.
- NCBI Sequence Archive. 2024. https://www.ncbi.nlm.nih.gov/datasets/ genome/?taxon=28450.
- Treangen TJ, Ondov BD, Koren S, Phillippy AM. The Harvest suite for rapid core-genome alignment and visualization of thousands of intraspecific microbial genomes. Genome Biol. 2014;15(11):524. https://doi.org/10.1186/ s13059-014-0524-x. PMID: 25410596; PMCID: PMC4262987.
- Chitlaru T, Cohen-Gihon I, Zvi A, Zaide G, Bilinsky G, Mamroud E. 2024. Figshare. https://doi.org/10.6084/m9.figshare.24961800.
- Letunic I, Bork P. Interactive Tree Of Life (iTOL): an online tool for phylogenetic tree display and annotation. Bioinformatics. 2007;23(1):127-8. https://doi. org/10.1093/bioinformatics/btl529. Epub 2006 Oct 18. PMID: 17050570.
- Dowling AJ. Novel gain of function approaches for vaccine candidate identification in Burkholderia pseudomallei. Front Cell Infect Microbiol. 2013;2:139. https://doi.org/10.3389/fcimb.2012.00139. PMID: 23316481; PMCID: PMC3540353.
- Balder R, Lipski S, Lazarus JJ, Grose W, Wooten RM, Hogan RJ, Woods DE, Lafontaine ER. Identification of Burkholderia mallei and Burkholderia pseudomallei adhesins for human respiratory epithelial cells. BMC Microbiol. 2010;10:250. https://doi.org/10.1186/1471-2180-10-250. PMID: 20920184; PMCID: PMC295563.
- Lazar Adler NR, Govan B, Cullinane M, Harper M, Adler B, Boyce JD. The molecular and cellular basis of pathogenesis in melioidosis: how does Burkholderia pseudomallei cause disease? FEMS Microbiol Rev. 2009;33(6):1079–99. https://doi.org/10.1111/j.1574-6976.2009.00189.x. Epub 2009 Aug 5. PMID: 19732156.
- Cohen-Gihon I, Chitlaru T. 2024. Figshare. https://doi.org/10.6084/ m9.figshare.25479535.

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